

# Computational study of the binding specificities of SH2 and SH3 domains

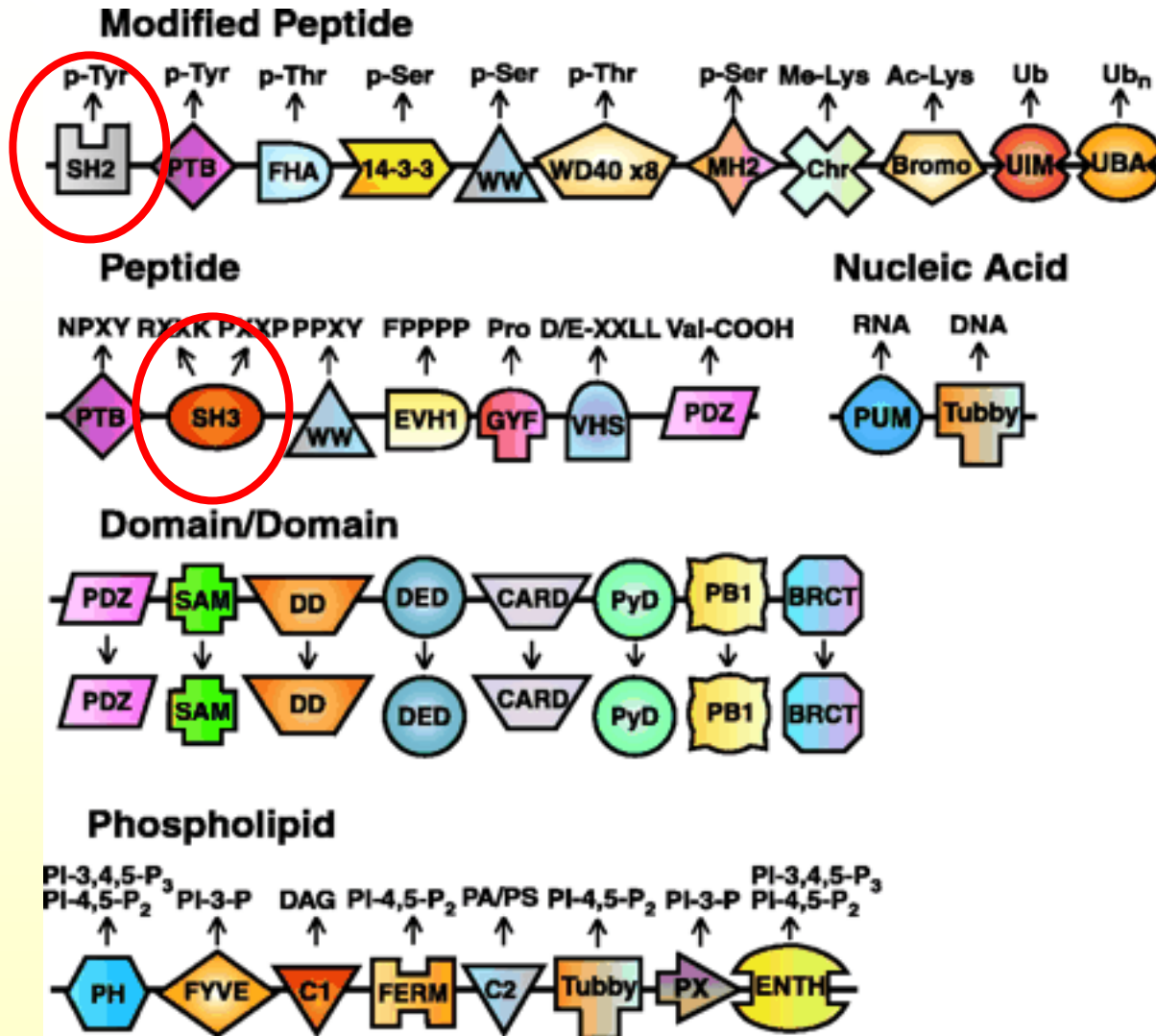
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# Modular design of protein-protein interactions:



Pawson and Nash, Science, 300, 445, 2003.

## Identification of protein-peptide interactions is challenging:

### 1. Issues with peptide library screening

A. still challenging to identify interacting partners given the binding motif.

B. may be biased by the artifacts of fixing peptides on the surface and/or the strong binding peptides not existing in the genome.

### 2. Issues with high throughput studies

Domain-peptide interactions are under-represented as the interactions are weak and transient.

### 3. Calculation of binding free energy for domain-peptide complex is time consuming.

## The first approach:

1. **Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
2. **Classify these peptides into binder and non-binder categories based on sequence and binding affinity.**
3. **Build a Hidden Markov Model (HMM) from binders and search the human proteome.**
4. **Remove false positives using conservation**
5. **Estimate the binding affinities of the top 100 candidates**
6. **Repeat 1-5.**

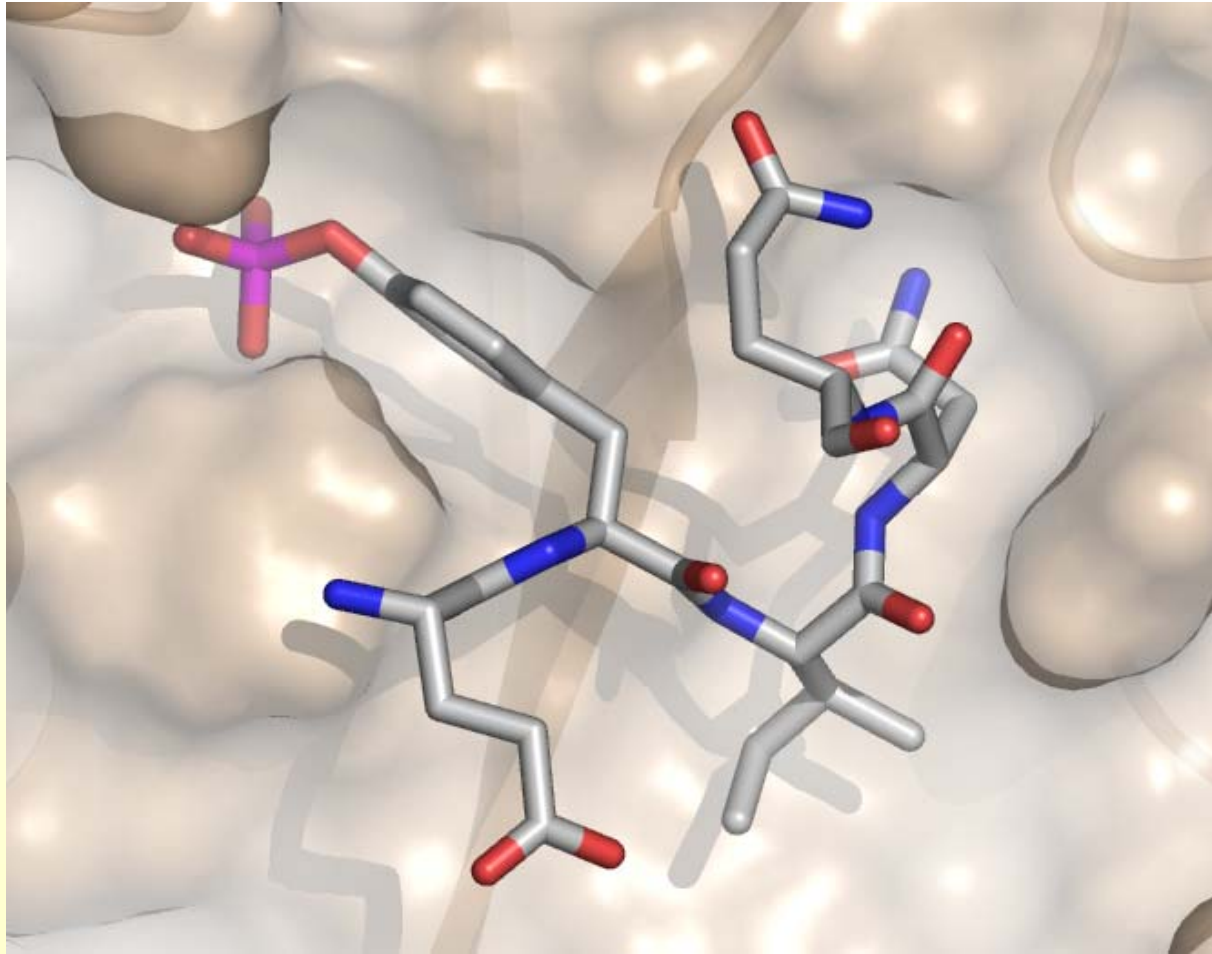
## Advantages of the first approach:

- 1. Do not require very accurate binding affinity calculation and only need to separate two distributions.**
- 2. Not biased by non-physiological strong binders  
All peptides present in the human proteome.**
- 3. Take the structural information into account .**

## The first approach:

1. **Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
  - A. **Model the complex structure from a known complex structure using a rotamer library;**
  - B. **Optimize the complex structure using AMBER;**
  - C. **Estimate binding free energy using MM/PBSA.**

## Known binder to the Grb2 SH2 domain



Sequence: Glu pTyr Ile Asn Gln

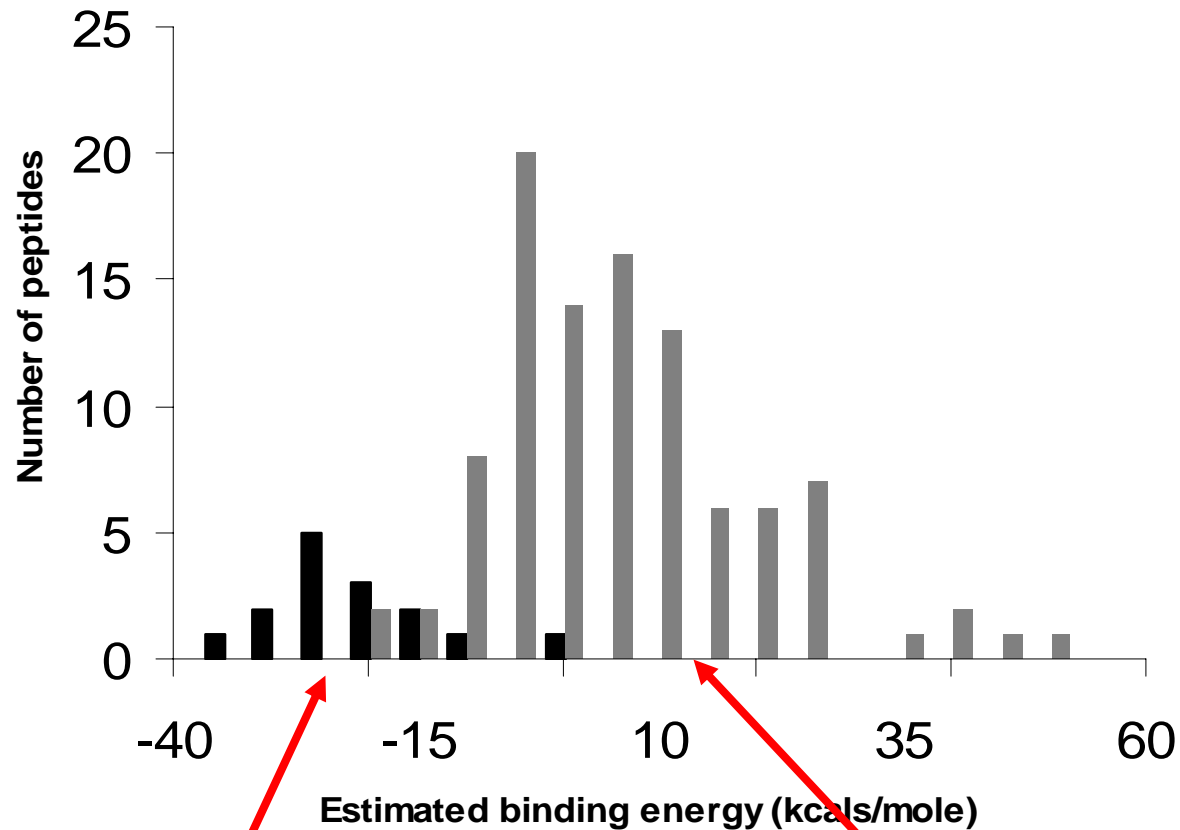
# The MM/PBSA (Molecular Mechanism/Poisson-Boltzmann Solvent Area) method

$$G = E_{MM} + G_{polar} + G_{non-polar} - TS$$

$$\begin{aligned}\Delta G_{bind} &= G_{complex} - G_{protein} - G_{ligand} \\ &= \Delta E_{MM} + \Delta G_{polar} + \Delta G_{nonpolar} - T\Delta S\end{aligned}$$



The binders and non-binders have distinct distributions.  
(Select 1400 peptides from the human proteome + 15 known binders)



15 known binders

100 randomly picked peptides

## The first approach:

1. **Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
2. **Classify these peptides into binder and non-binder categories based on sequence and binding affinity.**

Clusters created using both sequence and energy for the Grb2 dataset of peptides. Cluster “4” labeled as the binding cluster.

<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>Contents</b>
<b>357</b>	<b>13</b>	<b>262</b>	<b>118</b>	<b>425</b>	<b>225</b>	<b>Random Peptides</b>
<b>1</b>	<b>0</b>	<b>0</b>	<b>14</b>	<b>0</b>	<b>0</b>	<b>Known binders</b>

**Sequence only**

<b>1</b>	
<b>1400</b>	<b>Random</b>
<b>15</b>	<b>Known</b>

**Energy only**

<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	
<b>509</b>	<b>13</b>	<b>218</b>	<b>660</b>	<b>Random</b>
<b>14</b>	<b>0</b>	<b>0</b>	<b>1</b>	<b>Known</b>

## The first approach:

1. **Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
2. **Classify these peptides into binder and non-binder categories based on sequence and binding affinity.**
3. **Build a Hidden Markov Model (HMM) from binders and search the human proteome.**
  - **Using only the 15 known binders**
  - **Using peptides in the binding cluster**
  - **Using known binders plus peptide sequences from the nonbinding clusters**

## Grb2 SH2 binding sequence motifs (majority rule given by the HMMs)

Experimental motif from peptide array

...**Y**ΦNΦ...

Motif from known binders

dpe**Y**vNvts

Add binding cluster peptides

Add nonbinding cluster peptides

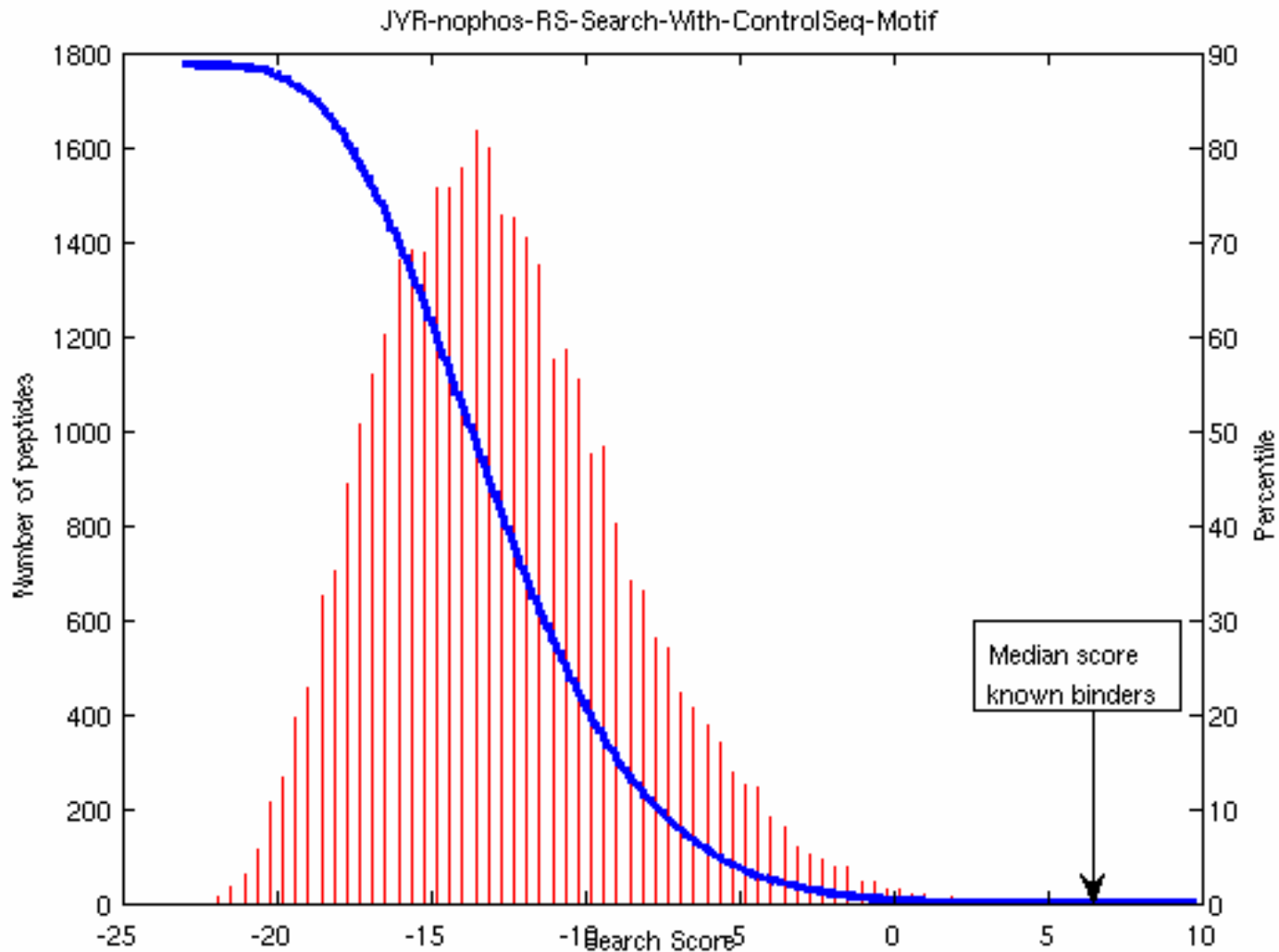
e.v**Y**vNl.l

...**Y**.lv.g

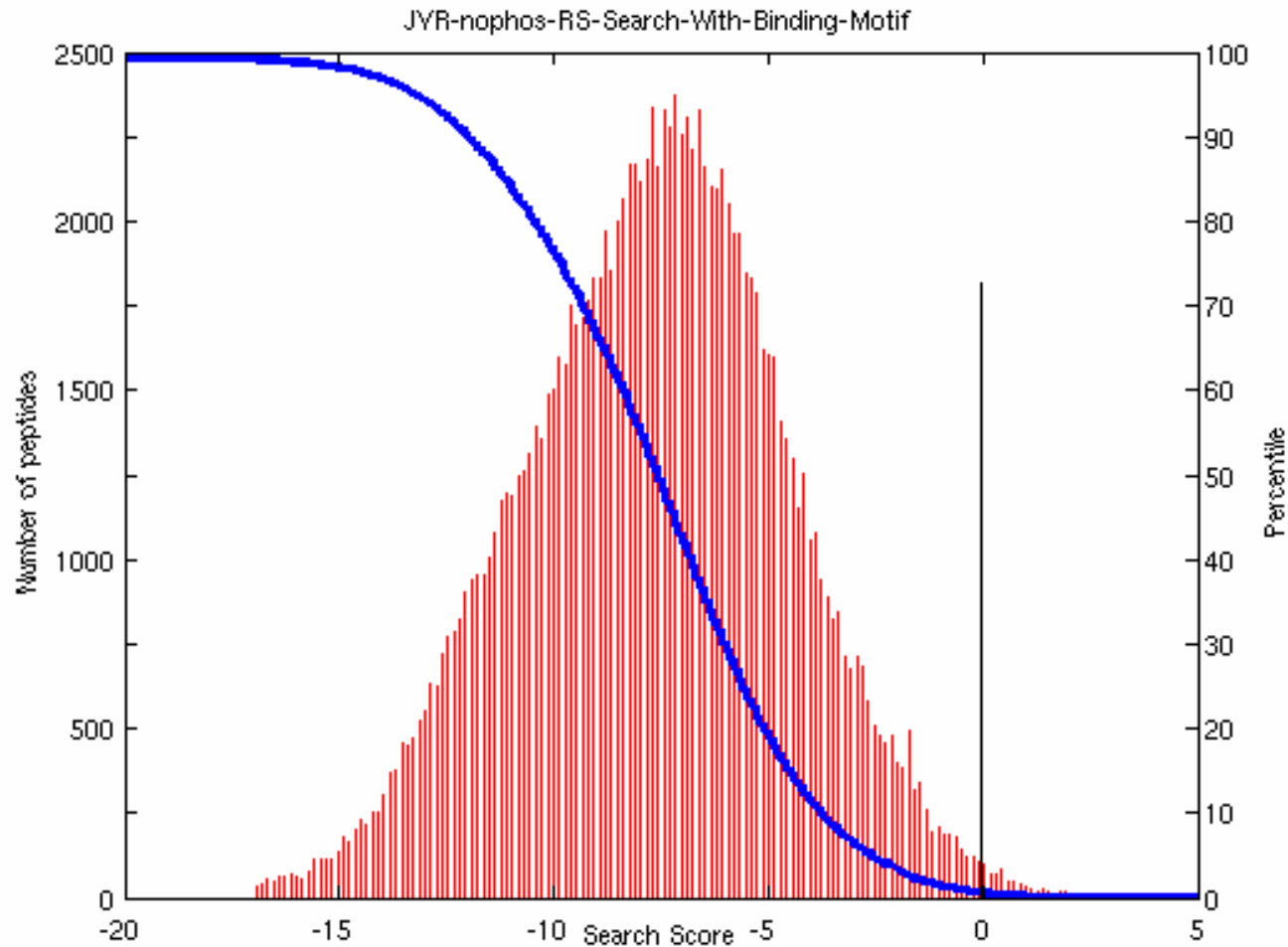
## Database screening:

- **Extract 174,604 peptides with xxxYxxxx sequences from the human proteins in SWISS-PROT**
- **Score all of the peptides using each of the HMMs**

# Known binders motif search

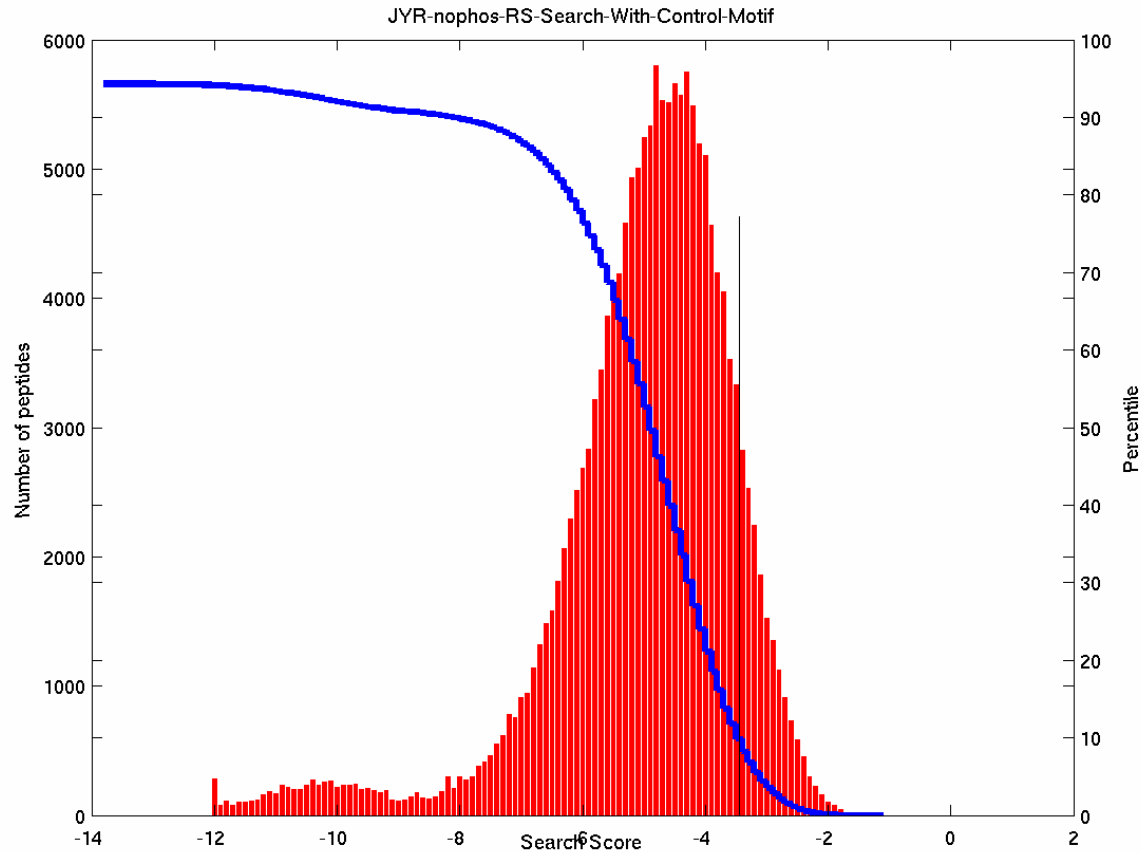


# Search with binding motif (HMM created with binding cluster peptide sequences)

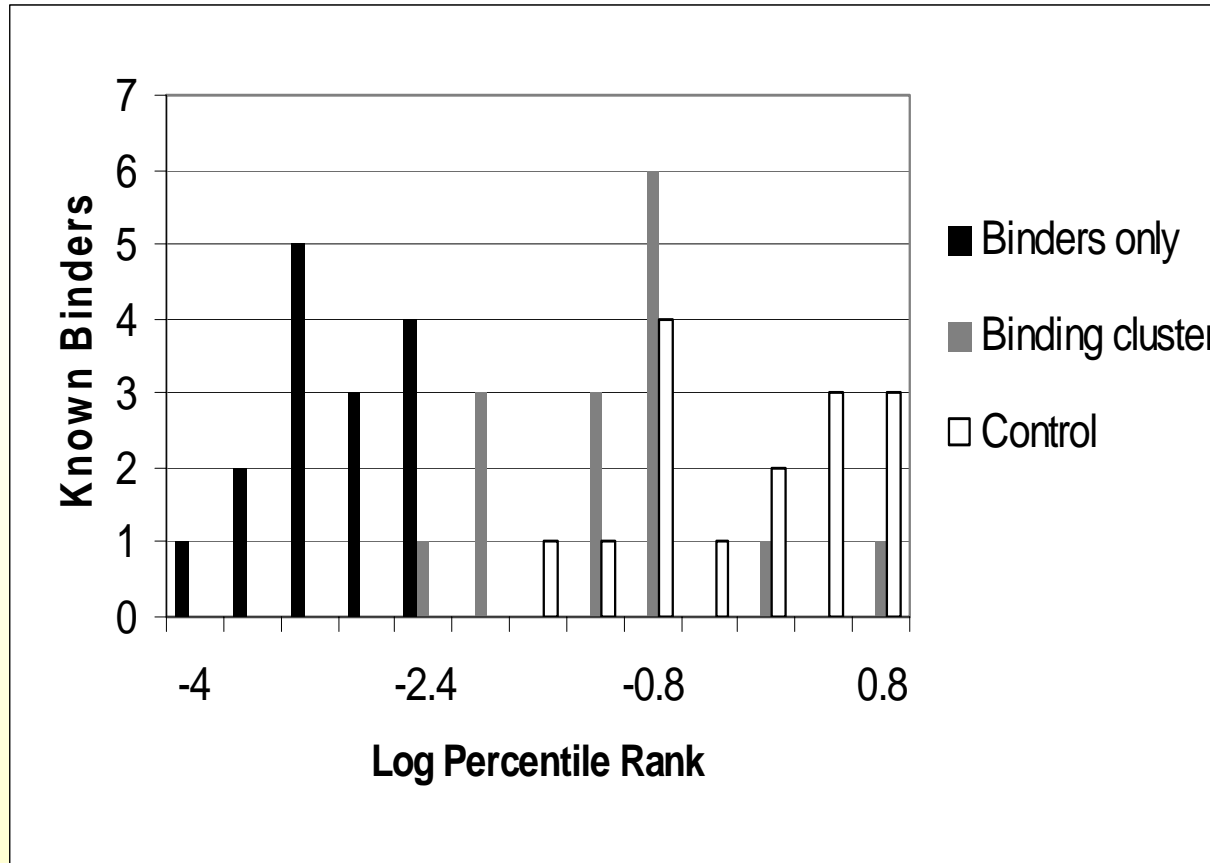




# HMM of known binders plus sequences from the non binding clusters



## Grb2 HMMs search results summary



**P-value of t-test comparing known binding ranks using binding cluster HMM and the control HMM = 0.032**

## Examine the top 100 hits of each search

- 1. Search with known binder HMM retrieved the known binders with little more.**
- 2. Search with binding cluster HMM retrieved many possible binders and one documented case (UFO, ranked 46 in our prediction but only 227 in Scansite output).**
- 3. Search with control HMM retrieved no viable candidates**

## The first approach:

- 1. Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
- 2. Classify these peptides into binder and non-binder categories based on sequence and binding affinity.**
- 3. Build a Hidden Markov Model (HMM) from binders and search the human proteome.**
- 4. Remove false positives using conservation**

# Examples of conserved peptides: UFO

Blast alignment for an example top hit: UFO protein

```
Human: 780 ELNPQDRPSFTELREDLENTLKALPPAQEPDEEILYVNMDEGGGYPEPPGAAGGADPPTQP 839
          ELNP+DRPSF ELREDLENTLKALPPAQEPDEEILYVNMDEGG + EP GAAGGADPPTQP
Mouse: 781 ELNPRDRPSFAELREDLENTLKALPPAQEPDEEILYVNMDEGGSHLEPRGAAGGADPPTQP 840
```

Comparison to the Grb2 binding motif

Grb2 binding motif \*->e.vYvNl.l<-\*

E +Y N+

UFO 1 EILYVNMDE 9

# Examples of conserved peptides: Nebulin

Blast alignment for an example top hit: Nebulin protein

```
Human : 2356 KFSSPVDMLGVVLAKKQCQLVSDVDYKNYLHQWTCLPDQNDVVQAKKVYELQSENLKSD 2415
          K++SPVDMLGVVLAKKQ LVSD DY+NYLHQWTCLPDQNDV+QAKKVYELQSEN+YKSD
Mouse : 241 KYTSPVDMLGVVLAKKQALVSDADYRNYLHQWTCLPDQNDVIQAKKVYELQSENMKSD 300
```

Comparison to the Grb2 binding motif

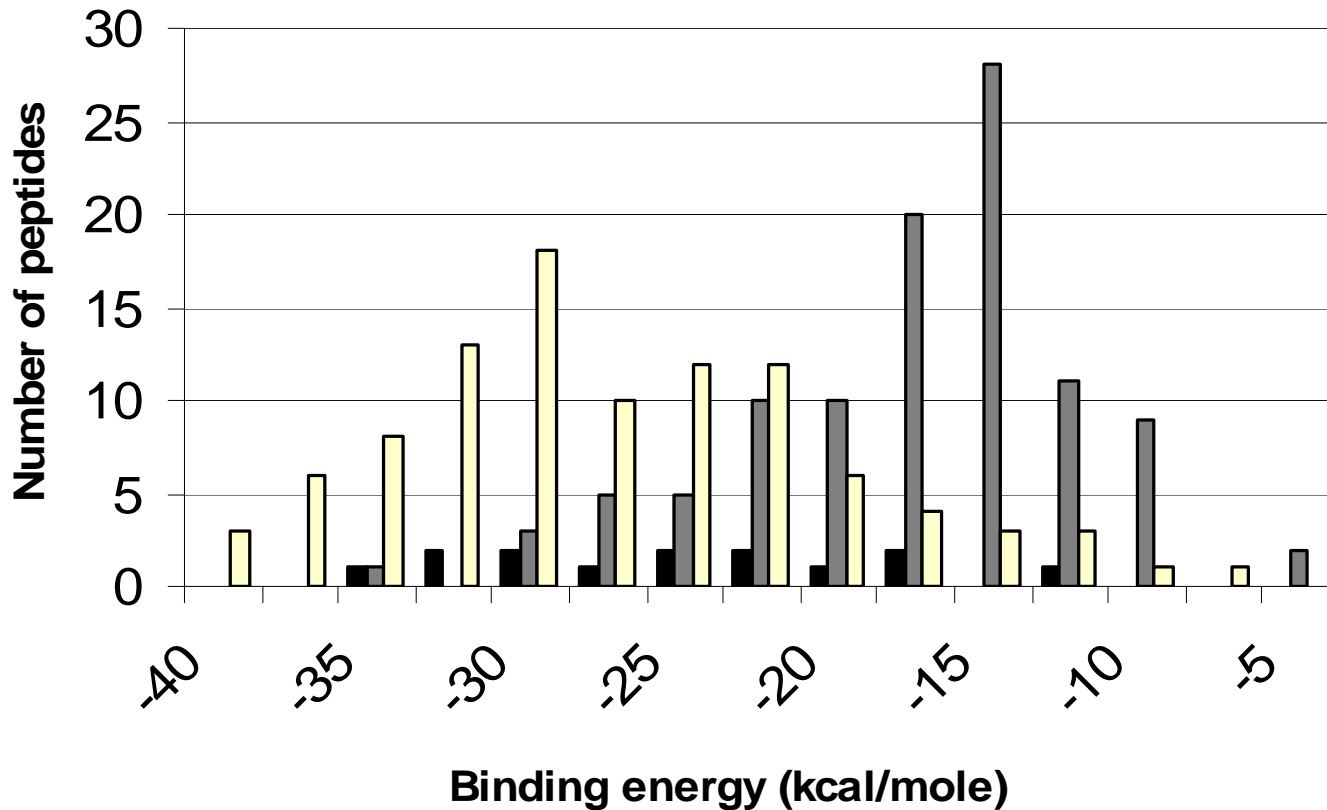
Score = **2.1**

```
Grb2 binding motif  *->e.vYvNl.l<-*
                    + +Y N+ +
Nebulin 2380      1   DaDYRNYlH      9
```

## The first approach:

1. **Roughly estimate the binding affinities of thousands of peptides selected from the human proteome.**
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4. **Remove false positives using conservation**
5. **Estimate the binding affinities of the top 100 candidates**

# The HMM captures both sequence and energy features



known binders (black), 100 random peptides in binding cluster (gray),  
top 100 predictions (white)



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6. **Repeat 1-5.**

# Evaluation of the top hits using sequence and energy (binding cluster is Cluster 5)

Clustering of top one hundred candidates plus original dataset

1	2	3	4	5	<-- assigned to cluster
270	13	510	503	104	Random peptides
0	0	0	1	14	Known binding
0	0	0	0	100	Top 100 from search

Cluster probabilities for the top ten candidates

Instance	Clus1	Clus2	Clus3	Clus4	Clus5	
0		0	0	0	0	1
1		0	0	0	0	1
2		0	0	0	0	1
3		0	0	0	0	1
4		0	0	0	0	1
5		0	0	0	0	1
6		0	0	0	0	1
7		0	0	0	0	1
8		0	0	0.00001	0	0.99999
9		0	0	0	0	1
10		0	0	0	0	1

## The second approach:

1. **Computational point mutation to generate a Position Specific Scoring Matrix (PSSM)**  
**Better consideration of conformational flexibility**
2. **Scan the database using this PSSM.**

## The second approach:

1. **Computational point mutation to generate a Position Specific Scoring Matrix (PSSM)**
  - A. **mutate each residue to other 19 amino acids**
  - B. **calculate the binding free energy using MM/PBSA**
  - C. **take the free energy difference between the mutated and the template peptides as the entry in the PSSM**
  
2. **Scan the database using this PSSM.**



Rank	Protein	Protein name	Start position	End position	Peptide	Score	Scansite Rank
1	RW1	RW1 protein [Fragment]	1521	1530	SPTPAS <u>P</u> S <u>P</u>	-4.06	Not in the top 2000
2	WASF4 (SCAR2)	Wiskott-Aldrich syndrome protein family member 4	475	484	PPPPSSPSFP	-3.59	Not in the top 2000
3	TREX1	Three prime repair exonuclease 1	107	116	GPPPTVPPPP	-3.38	1194
4	ACRO (ACR, ACRS)	Acrosin [Precursor]	344	353	PPPPSPPPP	-3.18	40
5	LRRN5 (GAC1)	Leucine-rich repeats neuronal protein 5 [Precursor]	22	31	VVPWHVPCPP	-2.94	Not in the top 2000
<b>6</b>	<b>SEM6A (SEMA6A)</b>	<b>Semaphorin 6A [Precursor]</b>	<b>791</b>	<b>800</b>	<b>MPPMGSPVIP</b>	<b>-2.89</b>	<b>Not in the top 2000</b>
7	HDAC4 (HD4)	Histone deacetylase 4	343	352	LPLYTSPSLP	-2.81	Not in the top 2000
<b>8</b>	<b>EVL (RNB6)</b>	<b>Ena/vasodilator stimulated phosphoprotein-like protein</b>	<b>185</b>	<b>194</b>	<b>PPPPPVPPPP</b>	<b>-2.65</b>	<b>83</b>
<b>9</b>	<b>WASF1 (WAVE1, WAVE-1)</b>	<b>Wiskott-Aldrich syndrome protein family member 1</b>	<b>347</b>	<b>356</b>	<b>TPPPPVPPPP</b>	<b>-2.65</b>	<b>132</b>
10	YLPM1 (ZAP3, ZAP113)	YLP motif containing protein 1	14	23	YPPPPVPPPP	-2.65	115

## Summary:

- 1. Computational approach and goal**
  - A. Identify binding motifs of modular domains**
  - B. Identify new physiological interacting partners**
- 2. Readiness of the application to study biological complex**
- 3. Bottleneck:**
  - A. Domain-peptide complex structures**
  - B. Experimental verification**
  - C. Nomenclature (gene names different in databases)**

## Acknowledgement

**Ken Chen**

**Han-Yu Chuang**

**Jie Liu**

**Tingjun Hou**

**Bill McLaughlin**

**Robert Shoemaker**

**Phil Bourne**

**Andy McCammon**

**Bing Ren**

**Yang Xu**

**Chanfeng Zhao (Illumina)**

**<http://wanglab.ucsd.edu>**