# Icosafiedral Reconstruction 

Wen I iang


PURDUE

## Icosafedral Viruses: Bacterioprages

 ssDNA MicroviridaeMicroviridae

dsDNA



Leviviridae

## dsRNA



Cystoviridae

Families and Genera of Viruses Infecting Vertebrates

## Icosafiedral VerteGrate Viruses


\(\left.\begin{array}{|l|}\hline dsRNA <br>
Reoviridae <br>
Orthoreovirus <br>
Orbivirus <br>
Coltivirus <br>
Rotavirus <br>

Aquareovirus\end{array}\right]\)| Birnaviridae |
| :---: |
| Aquabirnavirus |
| Avibirnavirus |



Rhabdoviridae
Lyssavirus Vesiculovirus Ephemerovirus Novirhabdovirus


Paramyxoviridae
Bornaviridae


| SsRNA (+) |  |  |
| :--- | :--- | :--- |
| Caliciviridae | Hepevirus | Nodaviridae <br> Betanodavirus |
| Togaviridae |  |  |

## Icosafedral Particles



ع15
$700 \AA$
HSV
$1250 \AA$

HBV
$300 \AA$


PDC 200Å

Mimivirus
5000Å

## Icosafiedral Symmetry

- 5, 3, 2 fold axes
- 6 five fold axes
- 10 three fold axes
- 15 two fold axes
- 60 fold symmetry in total

- Asymmetric unit: $1 / 3$ of the triangular face



## Image Processing

- In EMAN, icosahedral reconstruction is essentially treated the same as other symmetries
- However, one should pay special attention to the "large" problems
- Specialized methods exist for more efficient processing


## Image Processing

- Preprocessing
- Particle selection
- CTF fitting
- Build initial model
- Image refinement
- Orientation determination
- 3-D reconstruction


## Particle Selection

- Manual selection
- boxer
- Automated selection
- batchboxer
- ethan (for spherical particles)
- By Kivioja and Bamford et al.
- "Ring Filter"
- Only ONE parameter (Particle Size)
- Fast, 10-15 seconds / micrograph
- Tend to over-select


## Etran Example

Herpes
2.1 Å/pxiel 6662x8457 pixels 15 seconds in total
\$ time ethan.py jj0444-8bit.mrc 298600 jj0444.box jj0444.img
Opening file jj0444-8bit.mrc
Width: 6662
Height: 8457
Averaging 121 pixels
Filtering jj0444-8bit.mrc: **********
Total number of squares is 567
Number of peaks after sector test is 163
Number of peaks after first distance check is 81
Number of peaks after discarding too small ones is 80
Refining centers
74 particles found from jj0444-8bit.mrc
10.31s user 2.08s system $84 \%$ cpu 14.578 total

## Particle Selection

Suggested practice:

1. Automated selection using ethan
2. Manual screening using boxer

## CIF Fitting

Toreactifigtresolution, small defocuses (0.5-1.5 $\mu \mathrm{m}$ ) are oftenused for imaging large icosafedral particles


Saad et al, JSB 2001

## CIF Fitting

- Manual fitting
- ctfit
- Automated fitting
- fitctf
- fitctf.py
$I_{\text {inage }}(s)=I_{s}(s) C T F^{2}(s) e^{-2 s_{s}^{2}}+N(s)$
$\operatorname{ctf}(s)=A\left(\sqrt{1-Q^{2}} \sin (\gamma(s))+Q \cos (\gamma(s))\right)$
$\gamma(s)=-2 \pi\left(C_{s} \lambda^{3} s^{4}+Z \lambda s^{2} / 2\right)$
$N(s)=n_{3} e^{-n_{1} \sqrt{s}-n_{2}-\left(\pi n_{4} s\right)^{2} / 4}$
8 unknown parameters: $Z, B, A, Q$, $n 1, n 2, n 3, n 4$


## Algorithm:

constrained nonlinear optimization using a sequential quadratic programming (SQP) method)

$$
\min \left|I_{\text {image }}(s)-I_{s}(s) C T F^{2}(s) e^{-2 B s^{2}}-N(s)\right|^{2}
$$

subject to:

$$
\begin{aligned}
& I_{\text {image }}(s)>N(s) \\
& A>0 \\
& B \geq 0 \\
& 1 \geq Q \geq 0 \\
& \Delta Z<0 \text { (under focus) }
\end{aligned}
$$

Yang C. et al:
http://ncmi.bcm.edu/software/fitctf

## fitctf.py Examples



## CTF Fitting

Suggested practice:

1. Automated fitting using fitctf.py
2. Manual screening using ctfit

## Conventions: Orientation Origin



## MRC

- $\mathrm{X}, \mathrm{Y}, \mathrm{Z}$ axes along 2 fold sym axes for orientation ( $0,0,0$ )
- Euler: $Z \rightarrow Y^{\prime} \rightarrow Z^{\prime \prime}$


## EMAN

- Z along 5 fold sym axis
- $Y$ along 2 fold sym axis
- X near 3 fold sym axes for orientation (0,0,0)
- Euler: Z $\rightarrow$ X' $\rightarrow$ Z"


## Build Initial Model

- Classic method: self common-line
- Small number of particle with large defocus
- Self common-lines cluster for views near symmetry axes
- Standard EMAN method: starticos
- Find best 5-, 3-, 2-fold view particles
- Spherical particles cause problems
- New method: random model
- Build 3-D model from particles with randomly assigned orientations
- Relying on the wide convergence range to reach a correct structure


## 2- D Alignment Metfods

- Common-line method:
- Classic implementation: self common-line (global search to get coarse initial orientation) and cross common-line (local refinement) in tandem
- EMAN implementation: Cross common-line method for both global search and local refinement (EMAN crossCommonLineSearch.py command)
- Projection matching method (EMAN refine command)


## Cross Common-Line

Intersection of two planes in Fourier space

Z. Hong Zhou's thesis, p36

## Orientation $\rightarrow$ Common Line Locations

Fourier plane normal in 3D:

$$
\vec{n}_{r e f}=\vec{R}_{r e f} \cdot\left[\begin{array}{l}
0 \\
0 \\
1
\end{array}\right] \quad \vec{n}_{r a w}=\vec{R}_{r a w} \cdot\left[\begin{array}{l}
0 \\
0 \\
1
\end{array}\right]
$$

common line in 3D: $\quad \vec{C}=\vec{n}_{r a w} \times \vec{n}_{r e f}$
common line in 2D:

$$
\begin{array}{ll}
\vec{C}_{r e f}=\vec{R}_{r e f}{ }^{-1} \cdot \vec{C} & \vec{C}_{r a w}=\vec{R}_{r a w}{ }^{-1} \cdot \vec{C} \\
\alpha_{r e f}=\cos ^{-1}\left(\vec{C}_{r e f} \cdot\left[\begin{array}{l}
1 \\
0 \\
0
\end{array}\right]\right. & \alpha_{r a w}=\cos ^{-1}\left(\vec{C}_{r a w} \cdot\left[\begin{array}{l}
1 \\
0 \\
0
\end{array}\right]\right)
\end{array}
$$



## Common Line Searcf: Scoring Functions

search for the orientation and center parameters that minimize the mean phase differences among all the pairs of common lines

$$
R_{i}(\phi, \theta, \omega, x, y)=\frac{\sum_{j=1}^{N} \sum_{k=1}^{k_{\max }(j)} \sum_{s=s_{\min }}^{s_{\max }} f\left(\psi_{i}\left(s, x_{i}, y_{i}, \alpha_{i, j, k}\right), \psi_{j}\left(s, x_{j}, y_{j}, \alpha_{j, i, k}\right)\right) \times w\left(s, \alpha_{i, j, k}, \alpha_{j, i, k}\right)}{\left(s_{\max }-s_{\min }\right) \sum_{j=1}^{N} k_{\max }(j)}
$$

$i$ : particle image
$s$ : spatial frequency
$k$ : common-lines
$j$ : reference projections
$f($ : scoring function
$w()$ : weighting function

## Searcfi Strategy

- Classic implementation

1. center the particle by cross-correlation
2. self common-line to determine initial orientation by exhaustive searching orientation at $1^{\circ}$ step
3. cross common-line to locally refine the orientation and center by Gradient or Simplex.

Issues:

1. Self common-line is not very robust
2. Error propagation

## Searcfi Strategy

- Exhaustive cross common line search
- enumerate all centers and orientations in an asymmetric unit (5-level loops !)
- simple but really slow
- for 1 degree, 1 pixel step: $3 \times 10^{7}$ trials/particle


## Searcfi Strategy

- Multi-path Simulated Annealing
- Multiple randomly seeded SA processes
- Cross-communication among SA processes
- Global search for both orientation and center
- Accurate and fast
- $\sim 4 \times 10^{3}$ trials/particle.
- $>10^{4}$ times faster than exhaustive search


## Is The Best Solution Correct?

- Absolute score (residual) cutoff
- popular criterion
- needs different cutoff values for images at different defocuses
- how to pick a single number that suits all imaging conditions?


## Is The Best Solution Correct?

- Z score cutoff $\quad Z=\frac{x-\mu}{\sigma}$
- better, less sensitive to defocus
- unstable for very small defocus (<1 $\mu \mathrm{m}$ )



## Is The Best Solution Correct?

- Consistency criterion
- agreement among different methods (projection matching vs. common line)
- agreement among multiple runs of the global common line search
- more reliable than absolute value and Zscore cutoff
- bias toward "safer side"


## 3-D Reconstruction Metfods

- Fourier-Bessel Synthesis:
- Classic method for icosahedral particles
- Very efficient in computation
- Direct Fourier Inversion
- EMAN make3d command
- Needs lot of memory


## Fourier Bessel 3-D Reconstruction



- Efficient but incomplete use of data
- Anisotropic artifacts often seen

Icosafedral Reconstruction $\mathcal{Z l s i n g} E \mathcal{E A \mathcal { N }}$

## Two approaches

- Standard way using projection matching:
refine sym=icos <other options>
- Cross common-line method:
crossCommonLineSearch.py <options>


## Special Considerations

- Load projections to memory in batches instead all at once: add "projbatches=<n>" with $n>1$ to refine
- Use half maps instead of full map to start refine: use "proc3d <in> <out> tophalf" to generate top half map
- Use fewer CPUs for projection and class averaging step to avoid IO timeout errors. Use "proc=16,64" instead of "proc=64" option for refine


## Special Considerations

- Use parallel reconstruction instead of single CPU reconstruction. make3d is now parallelized using mpi.
mpiexec -n <\# cpu> make3d ...
- Map size limits: $512^{3}$ for 4 GB, $768^{3}$ for $8 G B, 900^{3}$ for 12GB
- Support mixed platform processing: one 64bit node with 12GB memory + many 32bit nodes with 4GB memory


## Icosafiedral Reconstruction $\rightarrow$ Non-

 Icosafiedral Reconstruction

## Bacteriop氏age Epsilon15


icosahedral reconstruction
(shell only)

asymmetric reconstruction (shell, portal, tail, genome)

## Asymmetric Reconstruction



## "Virus Anatomy"



$$
\begin{gathered}
\text { Icosafedral Reconstruction } \rightarrow \\
\mathcal{N} \text { (on-Icosafedral Reconstruction }
\end{gathered}
$$

refine
asymerfine.py


